

Ceragenins – a new weapon to fight multidrug resistant bacterial infections

Cerageniny – nowe perspektywy w zwalczaniu infekcji wywołanych przez wielooporne szczepy bakteryjne

Urszula Surel¹, Katarzyna Niemirowicz¹, Michal Marzec², Paul B. Savage³, Robert Bucki^{1,4}

¹Department of Microbiological and Nanobiomedical Engineering, Medical University of Białystok, Białystok, Poland
Head of Department: Prof. Adam Krętowski

²Department of Pathology and Laboratory Medicine, University of Pennsylvania, Philadelphia, PA, USA
Head of Department: Prof. David B. Roth

³Department of Chemistry and Biochemistry, Brigham Young University, Provo, UT, USA
Head of Department: Prof. Greg Burton

⁴Department of Pathophysiology and Infection Microbiology, Institute of Nursing and Obstetrics, Faculty of Health Sciences, Jan Kochanowski University, Kielce, Poland
Head of Department: Prof. Stanisław Głuszek MD, PhD

Studia Medyczne 2014; 30 (3): 207–213

Key words: microbiology, antibacterial activity, cationic lipids.

Słowa kluczowe: mikrobiologia, aktywność przeciwbakteryjna, lipidy kationowe.

Abstract

Growing antibiotic resistance among pathogenic microorganisms is one of the most challenging problems. Often, a single mutation in a bacterial cell leads to the formation of a new drug resistance mechanism. The ceragenins are a novel class of antibiotic, offering great promise in future treatment of infections. These cationic antimicrobial lipids are net positively charged cholic acid derivatives that are electrostatically attracted to the negatively charged membranes of bacteria, certain viruses, fungi, and protozoa. After membrane insertion, they interfere with membrane organisation, resulting in membrane dysfunction and cell death. This review focuses on the broad spectrum of antibacterial activity of ceragenins, and their potential to become a new group of antibiotics for prevention and treatment of infections, especially those caused by multidrug-resistant bacteria.

Streszczenie

Stale narastająca oporność bakterii na antybiotyki jest jednym z najtrudniejszych problemów. Często pojedyncza mutacja w komórce bakteryjnej prowadzi do powstania i rozwoju nowego mechanizmu, nadającego bakteriom oporność na antybiotyki. Cerageniny (pochodne kwasu cholowego) są analogami naturalnych kationowych peptydów przeciwbakteryjnych oferujących nowe możliwości w leczeniu infekcji bakteryjnych. Mają one dodatni ładunek powierzchniowy, dzięki czemu oddziałują elektrostycznie z negatywnie naładowaną powierzchnią bakterii, wirusów, grzybów i pierwotniaków. Po insercji w strukturę lipidową błony mikroorganizmów zaburzają jej funkcję, co w efekcie prowadzi do śmierci komórki. W niniejszej pracy przedstawiono szerokie spektrum aktywności przeciwdrobnoustrojowej ceragenin i ich potencjał w zwalczaniu infekcji, w szczególności powodowanych przez wielooporne szczepy bakteryjne.

Multidrug-resistance

The widespread inappropriate use of antibiotics is considered the major factor driving the increasing number of multidrug-resistant bacterial strains. Antibiotic treatment is very often prescribed as a preventative treatment and is given with disregard to the importance of the commensal microbiota that colonise the skin, gut, and mucosal surfaces of the human body [1]. According to the U.S. Center for Disease Control and Prevention (CDC), every year drug-resistant bacteria infect more than two million people nation-

wide, and a large percentage of those infections occur with involvement of multidrug-resistant bacteria. Additionally, some of those infections are acquired in health care facilities (health care-associated infections, HCAs). Multidrug-resistant pathogens usually cause infections in more vulnerable individuals, especially immunocompromised and immunosuppressed patients, and those with burn injuries, cancer, or genetic disorders such as cystic fibrosis (CF) or Down's syndrome [2, 3]. Drug resistance is considered the most important cause of expansion of tuberculosis

in the modern world. In the European Region of the World Health Organisation (WHO) a total of 15.7% of new and 45.3% of previously treated tuberculosis (TB) cases are estimated to be caused by multidrug-resistant tuberculosis (MDR-TB). Drug-resistant TB (XDR-TB) (resistance to fluoroquinolones and second-line injectables) has been reported extensively in 38 of the 53 countries of the region (72%) [4, 5]. In addition, there are an increasing number of reported infections caused by multidrug-resistant *Escherichia coli*, *Klebsiella pneumoniae*, *Staphylococcus aureus*, *Vibrio cholera*, and non-typhoid *Salmonella* in different African countries [6]. Some Asian countries have become epicentres of resistance, having seen rapid increases in the prevalence of antimicrobial resistance of major bacterial pathogens (MRSA, macrolide-resistant *Streptococcus pneumoniae*, and multidrug-resistant *Enterobacteriaceae*) with very high rates of HCAs [7, 8]. Latin America has a high rate of community-associated infections caused by multidrug-resistant *Enterobacteriaceae* relative to other world regions. Urinary tract infections (UTIs) by *E. coli*, and intra-abdominal infections (IAIs) by *E. coli* and *K. pneumoniae*, are characterised by high rates of resistance to trimethoprim/sulphamethoxazole, quinolones, and second-generation cephalosporins [9]. In response to the global public health threat posed by resistant pathogens a number of national and international actions and initiatives have been developed [10]. Although the most effective strategy to reduce the incidence of infections caused by multidrug-resistant bacteria has not yet been established, a multifaceted method is will probably be most effective, including actions aimed at optimising antibiotic use, increasing surveillance and infection control, and improving healthcare worker training and public education with regard to unanticipated consequences of antibiotic use [10]. Research should be focused on bringing new effective antibiotics, antibiotic-antibiotic combinations, and the development of adjuvants that either directly target resistance mechanisms ((such as inhibition of β -lactamase enzymes) or indirectly target resistance by interfering with bacterial signalling pathways (similarly to two-component systems (TCSs)) [11]. Design of new bactericidal molecules should be based on two fundamental principles. First, the new agents should target simple but fundamental properties of the bacteria, which would render resistance much more difficult to develop. Second, the antimicrobial agents should have anti-biofilm properties [12].

Ceragenins

Produced by shark *Squalus acanthias* and described in 1993, squalamine is considered to be the first natural representative of the ceragenin family (Figures 1 A and 1 B). It exhibits potent bactericidal activity against both Gram-negative and Gram-positive

bacteria. Furthermore, it is fungicidal by inducing osmotic lysis of the protozoa cell. The discovery of squalamine in the shark implicates a steroid molecule as a potential host-defence agent in vertebrates and provides insight into the chemical design of a family of broad-spectrum antibiotics [13]. In contrast to the sterol nature of fish squalamine, all mammals are equipped with cationic antibacterial peptides (CAPs) that represent the first line of defence against invasive pathogens [14, 15]. Physicochemical properties of squalamine and CAPs are similar because both are amphiphilic with net positive charge. Both are attractive candidates for clinical development of new antibiotics for three reasons: 1) a non-specific ability to induce dysfunction of the membranes of the pathogen (membrane permeabilisation and depolarisation), 2) speed of action, and 3) the difficulty of bacteria to develop a resistance mechanism [16–20].

The advantageous properties of squalamine and CAPs were used in the development of a new class of synthetic antibacterial molecules including ceragenins. Ceragenins are cholic acid derivatives [16] that are similar in antibacterial activity to condensed amino acid (derivatives of cholic acid marked with L-arginine), which was first synthesised in 1979 [21]. Like antibacterial peptides [22, 23], ceragenins display positive charges arranged on one face and hydrophobic residues on the other [16]. Ceragenins are also known as cationic steroid antibiotics (CSAs) and can be separated into two categories: polymyxin mimics, and squalamine and its mimics. Polymyxin mimics are characterised structurally by the attachment of three amine groups, via tethers, to a steroid nucleus. The second group consists of squalamine and its mimics, where the position of the polyamine and sulphate groups are reversed. Squalamine and its mimics can accept facially amphiphilic conformations in the presence of membrane molecules by passing the polyamine chain common to these compounds over the face of the steroid [24, 25]. CSA-13 is a lead compound from the ceragenin family, which is relatively simple to prepare and purify at a low cost [17, 19]. The broad spectrum of CSA-13 antibacterial activity includes activity against multidrug-resistant *P. aeruginosa* [26], vancomycin-resistant *S. aureus* [27] *H. pylori* [28], carbapenem-resistant *Acinetobacter baumannii* [29], and periodontopathic bacteria such as *Streptococcus mutans* and *Porphyromonas* species [30] (Table 1). Significant activity of CSA-13 against cariogenic and periodontopathic bacteria correlate with its ability to bind bacteria lipopolysaccharide and lipoteichoic acid linked to erythrocytes [30]. CSA-13 is also active against vaccinia virus (VV) [31] and *Trypanosoma cruzi* [32]. Although some forms of ceragenins are effective against both Gram-negative and Gram-positive bacteria, they are generally more potent against Gram-positive bacteria (Figures 1 C and 1 D). Surprisingly, it is

not the cell wall, but the high content of phosphatidylethanolamine in most Gram-negative bacteria that provide them with resistance [17]. Ceragenins with a hydrophobic chain are bactericidal at low concentrations and match the antibacterial activity of polymyxin B against Gram-positive bacteria [24]. Recently, antimicrobial nanoparticles were synthesised using ceragenins and they were introduced as multifunctional theranostics [33]. Different applications of ceragenins include contact lenses, hydrogels with an antibacterial innate immune function [34], polymeric coating applied to implanted devices to prevent perioperative device-related infections [35], thermally, chemically, and physically stable medical grade polydimethylsiloxane (PDMS) material to prevent biofilm formation [36], silicon [37], and gene delivery systems [38] (Figure 2). Similarly to cathelicidin-related antimicrobial peptides [15], ceragenins that mimic the hydrophobic and cationic morphology of cathelicidin have antiproliferative effects on the colon cancer-derived cell line HCT116. Addition of CSA-13 to a cell culture of HCT116 cells arrested cell growth, increasing the incidence of apoptosis detected by the binding of annexin V, and mitochondrial membrane depolarisation. More precisely, cell-cycle analysis showed that the CSA-13-treated wild-type and p53 null mutant HCT116 cell growth was arrested at the G1/S phase, indicating that CSA-13 affects the cell cycle through a p53-independent pathway. This finding suggests that the membrane-permeabilising capability is the common underlying mechanism for both the anticancer and antimicrobial effects of CSA-13 [39]. CSA-13 shows low toxicity in animal studies, supporting this compound's possible application in human treatment [40]. However, ceragenins and CAPs may be restricted to topical applications due to low activity in blood plasma [20]. Ceragenin molecules are advantageous over cationic amphipathic peptides due to their protease resistance. They also incorporate stably into membranes and have the unusual property of forming complexes with phospholipids [17].

Ceragenins in treatment of cystic fibrosis lung infections

Cystic fibrosis is an autosomal-recessive genetic disease caused by mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene of chromosome 7. Chronic lung infections caused in about 70% of CF adult patients by *P. aeruginosa* are the major cause of death in the course of CF lung disease. Treatment of lung infections to reduce inflammation and lung injury is of major importance in the management of CF. The CF individuals are extremely susceptible to bacterial infections of the respiratory tract due to very viscous, dehydrated sputum accumulating in the airways. Frequent and intensive antibiotic therapy is required to maintain lung function, to increase

Table 1. Susceptibility of selected bacteria strains to CSA-13 administration expressed as minimal inhibitory concentration (MIC)

Bacteria strain (*clinical isolate)	MIC [mg/l]	Ref.
<i>Staphylococcus aureus</i> MRSA	0.5	[20]
<i>Staphylococcus aureus</i> VISA	1	[20]
<i>Staphylococcus aureus</i> VRSA	1.1	[20]
<i>Staphylococcus aureus</i> ATCC 25923 VRSA	0.4	[18]
<i>Staphylococcus aureus</i> ATCC 25923	0.3	[18]
<i>Streptococcus salivarius</i> ATCC 13419	0.7	[44]
<i>Streptococcus mutans</i> ATCC 35668	0.7	[44]
<i>Staphylococcus epidermidis</i> *	0.35	[44]
<i>Streptococcus pneumoniae</i> *	0.35	[44]
<i>Streptococcus pyogenes</i> *	0.7	[44]
<i>Lactobacillus casei ssp. casei</i> ATCC 393	22.4	[44]
<i>Staphylococcus aureus</i> Xen 29	1.4	[44]
<i>Enterococcus faecalis</i> ATCC 29212	2.8	[44]
<i>Haemophilus influenzae</i> *	0.35	[44]
<i>Moraxella catarrhalis</i> ATCC 23246	1.4	[44]
<i>Helicobacter pylori</i> *	0.7	[44]
<i>Pseudomonas aeruginosa</i> Xen 5	5.6	[44]
<i>Pseudomonas aeruginosa</i>	2	[52]
<i>Pseudomonas aeruginosa</i> ATCC 27853	2	[18]
<i>Pseudomonas aeruginosa</i> 316*	4	[26]
<i>Pseudomonas aeruginosa</i> 711*	8	[26]
<i>Pseudomonas aeruginosa</i> 727*	1	[26]
<i>Pseudomonas aeruginosa</i> R1130	4	[26]
<i>Neisseria meningitidis</i> (B)	0.7	[44]
<i>Neisseria meningitidis</i> (C)	0.7	[44]
<i>Acinetobacter baumannii</i> ATCC 19606	3	[18]
<i>Acinetobacter baumannii</i>	1.6	[29]
<i>Pseudomonas cangingivalis</i>	3.2	[30]
<i>Pseudomonas circumdentaria</i>	0.8	[30]

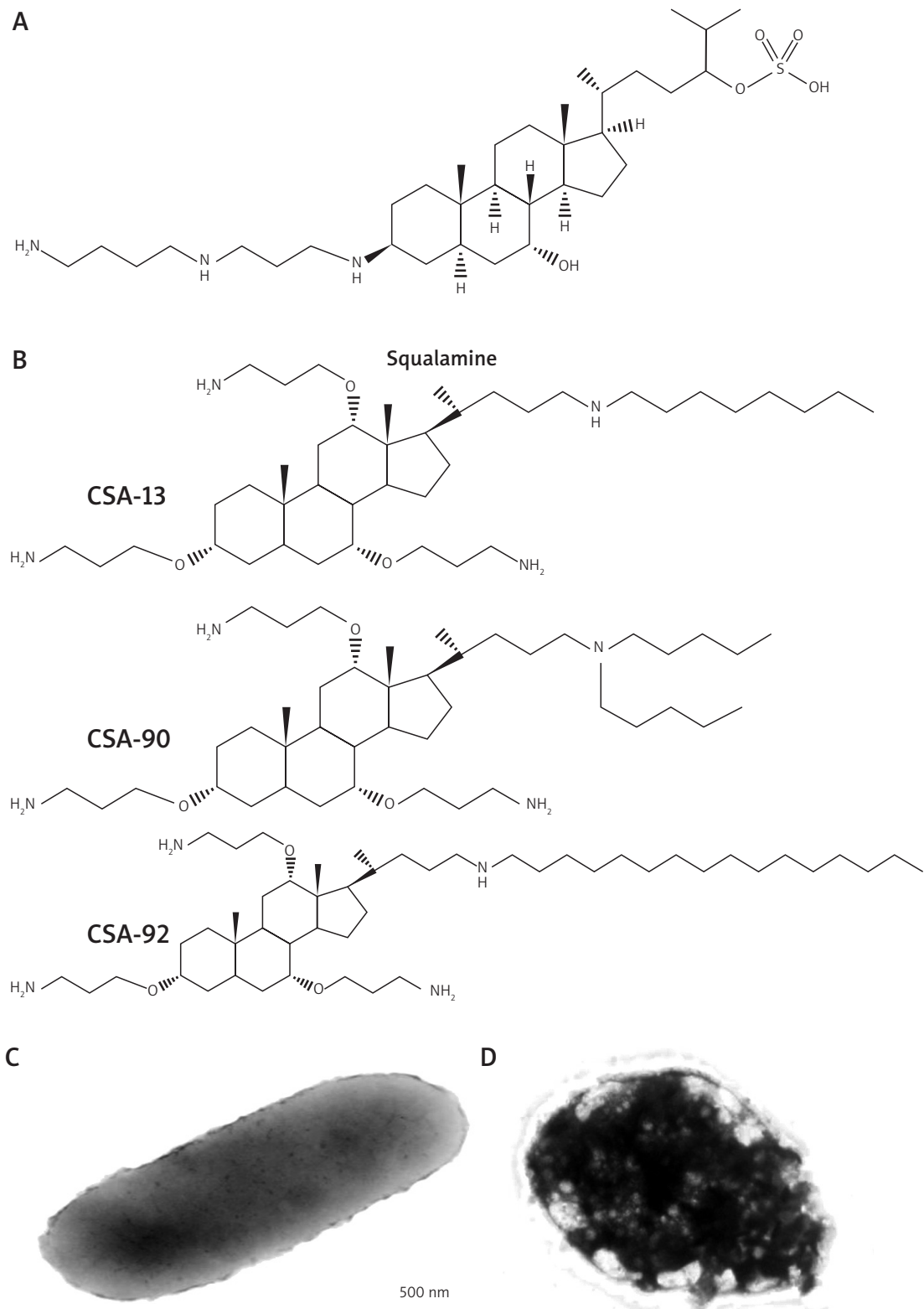


Figure 1. Squalamine: aminosterol molecules with potent broad spectrum of bactericidal activity isolated from tissues of the dogfish shark *Squalus acanthias* by Dr. Michael Zasloff [13] (panel A). Lead molecules of ceragenin family (panel B). EM image of *E. coli* cells before (panel C) and after treatment with CSA-13 for 1 h at 37°C (panel D)

quality of life, and to reduce exacerbations in infected patients [41]. Different studies suggest that ceragenins have strong potential for the development of new treatments for CF lung infections. The synergy of antibiotics with molecules contributing to innate immunity is an additional approach to fight multi-resistant bacteria [42]. In addition to *Pseudomonas aeruginosa*, other common pathogens of CF lung infections include: *Staphylococcus aureus*, *Haemophilus influenzae*, *Stenotrophomonas maltophilia*, and *Burkholderia species*. All are susceptible to ceragenin treatment *in vitro* [19, 43–45].

In CF airways, *P. aeruginosa* infection persists in biofilm form. Biofilm formation protects the aggregated, biopolymer-embedded bacteria from antibiotic treatments and host immunity [46]. Regardless of the morphology of the biofilm, its formation starts with the adhesion of bacterial cells. This process depends to some extent on the interaction overcoming any repulsive forces between microorganisms and components of the extracellular environment. Natural negatively charged biopolymers like DNA and F-actin released from host cells were recently identified as important factors stimulating *P. aeruginosa* biofilm growth [47] and are also a potential target to prevent biofilm formation [48, 49]. The antibacterial activity of ceragenins is not affected by DNA or F-actin, which are present in high concentration in cystic fibrosis airway sputum [43]. Combining ceragenins with classical antibiotics to fight resistant *P. aeruginosa* infections is a potential approach to this problem [50]. Bozkurt-Guzel *et al.* presented *in vitro* interactions of CSA-13 in combination with colistin, tobramycin, and ciprofloxacin against *P. aeruginosa* strains using a microbroth checkerboard. Their results showed synergistic interactions of CSA-13-colistin (54% of tested strains), whereas the least synergistic interactions were observed with the CSA-13-tobramycin (25% of tested strains). CSA-13-colistin is shown to be the most effective combination, and the frequency of synergistic interactions in this combination showed significant statistical differences from CSA-13-tobramycin and colistin-ciprofloxacin. This is the first study associating CSA-13 with colistin against *P. aeruginosa* strains isolated from CF patients. Nagat *et al.* showed that CSA-13 effectively kills ensconced cells within established biofilms, in addition to just on the surface [51]. A low concentration of CSA-13 inhibits the formation of a biofilm by *P. aeruginosa* through electrostatic interaction [12]. Therefore, CSA-13 has bactericidal activity against *P. aeruginosa* even in mature biofilms, and appears to be a good candidate for further investigations of the treatment involving biofilms of *P. aeruginosa* strains in CF patients [52].

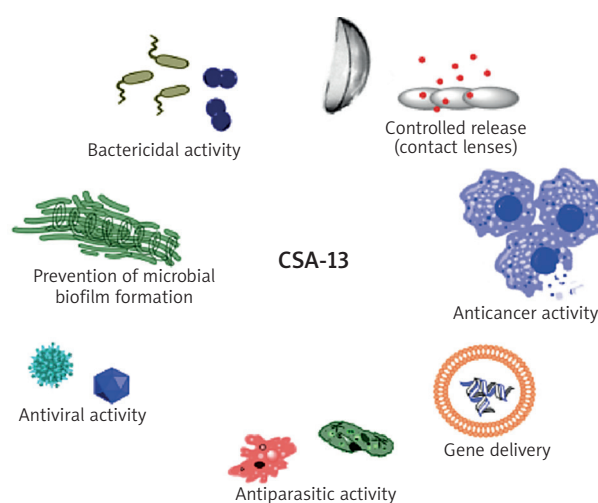


Figure 2. The various potential applications of ceragenin

Conclusions

Ceragenins are a promising class of molecules for the development of new treatments against infections caused by multidrug-resistant pathogens including resistant strains of *P. aeruginosa* within a biofilm.

References

1. Khanna S, Tosh PK. A clinician's primer on the role of the microbiome in human health and disease. *Mayo Clin Proc* 2014; 89: 107-14.
2. Shoham S, Shah PD. Impact of multidrug-resistant organisms on patients considered for lung transplantation. *Infect Dis Clin North Am* 2013; 27: 343-58.
3. Calfee DP. Multidrug-resistant organisms in dialysis patients. *Semin Dial* 2013; 26: 447-56.
4. Zignol M, Dara M, Dean AS, et al. Drug-resistant tuberculosis in the WHO European Region: an analysis of surveillance data. *Drug Resist Updat* 2013; 16: 108-15.
5. Zumla A, Raviglione M, Hafner R, von Reyn CF. Tuberculosis. *N Engl J Med* 2013; 368: 745-55.
6. Mshana SE, Matee M, Rweyemamu M. Antimicrobial resistance in human and animal pathogens in Zambia, Democratic Republic of Congo, Mozambique and Tanzania: an urgent need of a sustainable surveillance system. *Ann Clin Microbiol Antimicrob* 2013; 12: 28.
7. Kang CI, Song JH. Antimicrobial resistance in Asia: current epidemiology and clinical implications. *Infect Chemother* 2013; 45: 22-31.
8. Mathur P, Singh S. Multidrug resistance in bacteria: a serious patient safety challenge for India. *J Lab Physicians* 2013; 5: 5-10.
9. Salles MJ, Zurita J, Mejía C, et al. Resistant gram-negative infections in the outpatient setting in Latin America. *Epidemiol Infect* 2013; 141: 2459-72.
10. Paphitou NI. Antimicrobial resistance: action to combat the rising microbial challenges. *Int J Antimicrob Agents* 2013; 42 Suppl: S25-8.

11. Worthington RJ, Melander C. Combination approaches to combat multidrug-resistant bacteria. *Trends Biotechnol* 2013; 31: 177-84.
12. Nagant C, Feng Y, Lucas B, et al. Effect of a low concentration of a cationic steroid antibiotic (CSA-13) on the formation of a biofilm by *Pseudomonas aeruginosa*. *J Appl Microbiol* 2011; 111: 763-72.
13. Moore KS, Wehrli S, Roder H, et al. Squalamine: an aminosterol antibiotic from the shark. *Proc Natl Acad Sci U S A* 1993; 90: 1354-8.
14. Zasloff M. Magainins, a class of antimicrobial peptides from *Xenopus* skin: isolation, characterization of two active forms, and partial cDNA sequence of a precursor. *Proc Natl Acad Sci U S A* 1987; 84: 5449-53.
15. Zasloff M. Antimicrobial peptides in health and disease. *N Engl J Med* 2002; 347: 1199-200.
16. Ding B, Taotofa U, Orsak T, et al. Synthesis and characterization of peptide-cationic steroid antibiotic conjugates. *Org Lett* 2004; 6: 3433-6.
17. Epanand RM, Epanand RF, Savage PB. Ceragenins (cationic steroid compounds), a novel class of antimicrobial agents. *Drug News Perspect* 2008; 21: 307-11.
18. Pollard JE, Snarr J, Chaudhary V, et al. In vitro evaluation of the potential for resistance development to ceragenin CSA-13. *J Antimicrob Chemother* 2012; 67: 2665-72.
19. Lai XZ, Feng Y, Pollard J, et al. Ceragenins: cholic acid-based mimics of antimicrobial peptides. *Acc Chem Res* 2008; 41: 1233-40.
20. Van Bambeke F, Mingeot-Leclercq MP, Struelens MJ, Tulkens PM. The bacterial envelope as a target for novel anti-MRSA antibiotics. *Trends Pharmacol Sci* 2008; 29: 124-34.
21. Bellini AM, Vertuani G, Quaglio MP, Cavazzini G. Bile acid derivatives with antimicrobial activity. *Farmaco Sci* 1979; 34: 967-78.
22. Zanetti M, Gennaro R, Scocchi M, Skerlavaj B. Structure and biology of cathelicidins. *Adv Exp Med Biol* 2000; 479: 203-18.
23. Bucki R, Pastore JJ, Randhawa P, et al. Antibacterial activities of rhodamine B-conjugated gelsolin-derived peptides compared to those of the antimicrobial peptides cathelicidin LL37, magainin II, and melittin. *Antimicrob Agents Chemother* 2004; 48: 1526-33.
24. Savage PB, Li C, Taotofa U, et al. Antibacterial properties of cationic steroid antibiotics. *FEMS Microbiol Lett* 2002; 217: 1-7.
25. Deng G, Dewa T, Regen SL. A synthetic ionophore that recognizes negatively charged phospholipid membranes. *J Am Chem Soc* 1996; 118: 8975-6.
26. Chin JN, Jones RN, Sader HS, et al. Potential synergy activity of the novel ceragenin, CSA-13, against clinical isolates of *Pseudomonas aeruginosa*, including multidrug-resistant *P. aeruginosa*. *J Antimicrob Chemother* 2008; 61: 365-70.
27. Chin JN, Rybak MJ, Cheung CM, Savage PB. Antimicrobial activities of ceragenins against clinical isolates of resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2007; 51: 1268-73.
28. Leszczynska K, Namiot A, Fein DE, et al. Bactericidal activities of the cationic steroid CSA-13 and the cathelicidin peptide LL-37 against *Helicobacter pylori* in simulated gastric juice. *BMC Microbiol* 2009; 9: 187.
29. Bozkurt-Guzel C, Savage PB, Akcali A, Ozbek-Celik B. Potential synergy activity of the novel ceragenin, CSA-13, against carbapenem-resistant *Acinetobacter baumannii* strains isolated from bacteremia patients. *Biomed Res Int* 2014; 2014: 710273.
30. Isogai E, Isogai H, Takahashi K, et al. Ceragenin CSA-13 exhibits antimicrobial activity against cariogenic and periodontopathic bacteria. *Oral Microbiol Immunol* 2009; 24: 170-2.
31. Howell MD, Streib JE, Kim BE, et al. Ceragenins: a class of antiviral compounds to treat orthopox infections. *J Invest Dermatol* 2009; 129: 2668-75.
32. Lara D, Feng Y, Bader J, et al. Anti-trypanosomatid activity of ceragenins. *J Parasitol* 2010; 96: 638-42.
33. Hoppens MA, Wheeler ZE, Qureshi AT, et al. Maghemite, silver, ceragenin conjugate particles for selective binding and contrast of bacteria. *J Colloid Interface Sci* 2014; 413: 167-74.
34. Gu X, Jennings JD, Snarr J, et al. Optimization of ceragenins for prevention of bacterial colonization of hydrogel contact lenses. *Invest Ophthalmol Vis Sci* 2013; 54: 6217-23.
35. Sinclair KD, Pham TX, Williams DL, et al. Model development for determining the efficacy of a combination coating for the prevention of perioperative device related infections: a pilot study. *J Biomed Mater Res B Appl Biomater* 2013; 101: 1143-53.
36. Williams DL, Sinclair KD, Jeyapalina S, Bloebaum RD. Characterization of a novel active release coating to prevent biofilm implant-related infections. *J Biomed Mater Res B Appl Biomater* 2013; 101: 1078-89.
37. Sinclair KD, Pham TX, Farnsworth RW, et al. Development of a broad spectrum polymer-released antimicrobial coating for the prevention of resistant strain bacterial infections. *J Biomed Mater Res A* 2012; 100: 2732-8.
38. Kichler A, Leborgne C, Savage PB, Danos O. Cationic steroid antibiotics demonstrate DNA delivery properties. *J Control Release* 2005; 107: 174-82.
39. Kuroda K, Fukuda T, Okumura K, et al. Ceragenin CSA-13 induces cell cycle arrest and antiproliferative effects in wild-type and p53 null mutant HCT116 colon cancer cells. *Anticancer Drugs* 2013; 24: 826-34.
40. Saha S, Savage PB, Bal M. Enhancement of the efficacy of erythromycin in multiple antibiotic-resistant gram-negative bacterial pathogens. *J Appl Microbiol* 2008; 105: 822-8.
41. Dhooghe B, Noël S, Huaux F, Leal T. Lung inflammation in cystic fibrosis: pathogenesis and novel therapies. *Clin Biochem* 2014; 47: 539-46.
42. Rosenthal KL. Tweaking innate immunity: the promise of innate immunologicals as anti-infectives. *Can J Infect Dis Med Microbiol* 2006; 17: 307-14.
43. Bucki R, Sostarecz AG, Byfield FJ, et al. Resistance of the antibacterial agent ceragenin CSA-13 to inactivation by DNA or F-actin and its activity in cystic fibrosis sputum. *J Antimicrob Chemother* 2007; 60: 535-45.
44. Leszczynska K, Namiot D, Byfield FJ, et al. Antibacterial activity of the human host defence peptide LL-37 and selected synthetic cationic lipids against bacteria associated with oral and upper respiratory tract infections. *J Antimicrob Chemother* 2013; 68: 610-8.
45. Rodríguez-Rojas A, Oliver A, Blázquez J. Intrinsic and environmental mutagenesis drive diversification and persistence of *Pseudomonas aeruginosa* in chronic lung infections. *J Infect Dis* 2012; 205: 121-7.

46. Alhede M, Bjarnsholt T, Givskov M. *Pseudomonas aeruginosa* biofilms: mechanisms of immune evasion. *Adv Appl Microbiol* 2014; 86: 1-40.
47. Walker TS, Tomlin KL, Worthen GS, et al. Enhanced *Pseudomonas aeruginosa* biofilm development mediated by human neutrophils. *Infect Immun* 2005; 73: 3693-701.
48. Parks QM, Young RL, Poch KR, et al. Neutrophil enhancement of *Pseudomonas aeruginosa* biofilm development: human F-actin and DNA as targets for therapy. *J Med Microbiol* 2009; 58: 492-502.
49. Moreau-Marquis S, Stanton BA, O'Toole GA. *Pseudomonas aeruginosa* biofilm formation in the cystic fibrosis airway. *Pulm Pharmacol Ther* 2008; 21: 595-9.
50. Döring G, Conway SP, Heijerman HG, et al. Antibiotic therapy against *Pseudomonas aeruginosa* in cystic fibrosis: a European consensus. *Eur Respir J* 2000; 16: 749-67.
51. Nagant C, Pitts B, Stewart PS, et al. Study of the effect of antimicrobial peptide mimic, CSA-13, on an established biofilm formed by *Pseudomonas aeruginosa*. *Microbiolgyopen* 2013; 2: 318-25.
52. Bozkurt-Guzel C, Savage PB, Gerceker AA. In vitro activities of the novel ceragenin CSA-13, alone or in combination with colistin, tobramycin, and ciprofloxacin, against *Pseudomonas aeruginosa* strains isolated from cystic fibrosis patients. *Chemotherapy* 2011; 57: 505-10

Address for correspondence:

Prof. **Robert Bucki**

Department of Pathophysiology
and Infection Microbiology
Faculty of Health Sciences
Jan Kochanowski University
ul. IX Wieków 19, Kielce, Poland
Fax: +48 85 748 54 83
E-mail: mikro.nano@umb.edu.pl