

# ALLICIN HAS SIGNIFICANT EFFECT ON AUTOIMMUNE ANTI-ISLET CELL ANTIBODIES IN TYPE 1 DIABETIC RATS

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The research purpose was to experimentally investigate the effect of allicin administration on the levels of main type 1 diabetes (IDDM) autoantibodies which are anti-islet cell antibodies (ICA) with an attempt to find a relation between this immunological effect and histological and/or biochemical findings. We have evaluated, with the help of ELISA kits, the levels of ICA and serum insulin in male Sprague-Dawley rats with Streptozotocin-induced IDDM in addition to pancreatic histological findings. The four groups (6 rats each) under study received or not different intraperitoneal doses of allicin for a period of 30 days. Daily intraperitoneal administration of allicin (either at as low dose of 8 mg/kg or high dose of 16 mg/kg) for up to 30 days to type 1 diabetic rats effectively reduces levels of anti-islet cell antibodies and in addition, reduced the level of insulin due to damaged Langerhans islet cell was significantly increased in the serum due to a repairing tissue process in pancreatic tissues. These experimental results suggest that allicin treatment has a therapeutic protective effect against autoimmune reactions occurring in IDDM. The data may provide new strategies for using allicin to be recommended as an excellent candidate in the clinical management, control, and prevention of IDDM.

**Key words:** allicin, garlic, type 1 diabetes mellitus, anti-islet cell antibody, serum insulin.

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## Introduction

Type 1 diabetes mellitus or insulin-dependent diabetes mellitus (IDDM) is an autoimmune disease process in which pancreatic islet  $\beta$ -cells are targeted for destruction by an aberrant host immune system [1]. This process involves both the cellular and humoral branches of the immune system, with the generation of islet-specific T-cell reactivity, as well as autoantibodies directed against islet cell antigens [1].

The disease process in IDDM is primarily caused by the destruction of pancreatic  $\beta$  cells that is thought to result mainly from the action of T-lymphocytes; the key players in autoimmune disease development [2]. Meanwhile, in subjects newly diagnosed with IDDM, up to 90% have autoantibodies to islet cell antibodies ICA [2]. These autoantibodies appear during the pre-

clinical period of  $\beta$ -cell destruction before the clinical manifestation of diabetes [2]. The hallmark of the autoimmunity of IDDM is the presence of circulating ICA autoantibodies, they are thought to signal a T-cell mediated immune response which sets the stage for  $\beta$  cell destruction [2]. However, multiple environmental and genetic factors make the immune cells, particularly T lymphocytes, to invade islet  $\beta$  cells and cause pancreatic inflammation [3].

Experimental studies on animals, especially rats, showed that these animals develop a form of autoimmune diabetes that resembles human IDDM [4]. Studies with an animal model have established that islet-infiltrating cell-reactive T-cells are the major effectors of  $\beta$ -cell damage. However, other immune system cells are also crucial in the disease development. Among these cells, B-cells are essential in the onset and progression

of type 1 diabetes [5, 6], and although it is not fully understood when and how these cells participate in IDDM, it is known that they produce ICA autoantibodies against many  $\beta$ -cell autoantigens [6] and act as antigen-presenting cells [7]. On the other hand, the production of specific ICA autoantibodies directly correlates with the progression of IDDM in both humans and laboratory animals [7].

Due to the increasing worldwide prevalence and financial burden of diabetes, it has become increasingly important to find pharmacological remedies to alleviate the symptoms and complications of these conditions. In particular, the use of natural remedies such as garlic and its major component allicin, has become popular as both preventative and treatment alternatives [1].

Allicin is a major component of garlic (*Allium sativum*) and a precursor of many secondary products formed in aged garlic and crushed garlic preparations. The antimicrobial [8, 9], antitumor [10-12], antifungal [13], and antigenotoxic [14], activities of allicin have been reported. Meanwhile, various researches have indicated that garlic and allicin have inhibitory immunomodulatory action [15], they modulate immune responses, enhance humoral immunity and minimize immunological stress, and thus affect growth performance most positively [15]. Studies demonstrated that garlic enhances natural killer (NK) activity and T-lymphocyte proliferation [15, 16].

Many studies have examined the hypoglycemic effect of allicin in both types of DM [17-20], but till now the mechanism has not been discussed with regard to IDDM while the probable mechanism underlying garlic and allicin hypoglycemic effects in type 2 diabetes most likely is increased insulin secretion and sensitivity [21, 22].

To date, there have been no successful treatment interventions that have been found to delay the onset of type 1 diabetes. Thus, our present study was carried out in UiTM Malaysia, for the first time according to our knowledge to investigate the effect of allicin administration on the levels of main IDDM autoantibodies which are anti-islet cell antibodies with an attempt to find a relation between this immunological effect and histological and/or biochemical findings.

## Material and methods

### Experimental animals

Twenty four male Sprague-Dawley rats with an average weight of 150-250 g and an average age of 12-16 weeks were used throughout the experiment, obtained from Nano Life Quest Company. The rats were acclimatized for a period of 21 days. Standard environmental conditions such as temperature (20-22°C), relative humidity (45-55%) and 12 hrs dark/light cy-

cles were maintained. The animals were fed daily with rodent pellet diet and tap water *ad libitum* under strict hygienic conditions.

Ethical clearance for performing the experiment on animals was approved by the Animal Care and Use Committee (ACUC), Faculty of Medicine, Universiti Teknologi MARA (UiTM) Malaysia that conforms to the Guide for the Care and Use of Laboratory Animals [20] and all efforts were made to minimize animal suffering and the number of animals used.

### Chemicals

Streptozotocin (STZ) used in the present study was purchased from Nano Life Quest Company; Allicin (2-propene-1-sulfinothioic acid S-2-propenyl ester; thio-2-propene-1-sulfinic acid S-allyl ester) was purchased from Nano Life Quest Company. The allicin was administered once a day by intraperitoneal injection (i.p.) at a dose of 8 mg/kg and 16 mg/kg for 30 days.

### Induction of type 1 diabetes mellitus and treatment of rats

A single injection of STZ is widely used to generate a rat model of type 1 diabetes, which results from the selective toxicity of STZ towards the insulin-producing  $\beta$ -cells in pancreatic islets [23, 24]. IDDM was induced in an overnight fasted animal group by intraperitoneal injection with a single dose of STZ (65 mg/kg body weight) (Sigma). This dose of STZ lies within the range used in most of studies to produce IDDM, in which blood glucose levels are 3-4 times higher than normal, by causing a substantial depletion of pancreatic insulin [25]. Streptozotocin was dissolved in sodium citrate buffer solution (PH 4.5) immediately before use. The development of IDDM was confirmed by the presence of hyperglycemia with blood glucose above 13.9 mmol/l (250 mg/dl), which lasted for at least three days. The rats were divided into four groups comprising 6 rats each. Group A (GA; control group, rats were injected with an equal volume of vehicle (citrate buffer, 65 mg/kg body weight)); Group B (GB; untreated STZ-diabetic rats); Group C (GC; STZ-diabetic rats treated with 8 mg/kg, i.p., allicin); Group D (GD; STZ-diabetic rats treated with 16 mg/kg, i.p., allicin).

The treatment by allicin was started for a period of 30 days. During this period, all animals had free access to standard diet and water until 6 pm. None of the rats was treated with insulin at any time during the experiment. Animals were sacrificed on the 30<sup>th</sup> day of experiment immediately after measuring blood glucose [24]. Blood glucose levels were tested every morning (at 8 am). Blood was collected from the tail of fasting (14 h) animals. A drop of blood was used for the blood glucose test with the help of a One Touch Glucometer GX.

### Laboratory tests

On the last day (30<sup>th</sup> day) and after completion of the experimental protocols, blood samples were collected from overnight fasting rats by sacrificing each diabetic and control rats. The animals were anesthetized in a chamber containing diethyl ether. A cardiac puncture was done using a heparin syringe and blood was collected into a heparin containing container. Immediately after collection, 2.0 ml of blood was transferred into a fresh tube and centrifuged at 3000 rpm for 10 minutes. The serum was collected and stored at -80°C until serological analysis.

Serum was assayed for anti-islet cell antibodies (ICA) and serum insulin using enzyme-linked immunosorbent assay (ELISA) using commercially available kits (USCNK, CHINA). Also pancreatic tissues were collected for histological examination.

### Summary of histopathological procedures

Pancreatic tissues [26] were harvested from the animals and they were fixed in 10% neutral formalin solution, embedded in paraffin, and then stained with hematoxylin and eosin (HE). The preparations were evaluated by means of a bright-field microscope and photographed (Optiphot 2; Nikon, Tokyo, Japan).

### Statistical analysis

The data are expressed as mean ± SE with 'n' referring to the number of rats used. Two-way analysis of variance (ANOVA) was carried out using SPSS 16 software to assess the overall effects and interaction of treatment and time on parameters and followed by repeated one-way analysis of variance (ANOVA) with a post hoc least significant difference (LSD) test to determine the effect of treatment on differences among means when the analysis of variance indicated a significant result. P < 0.05 was taken to indicate significance.

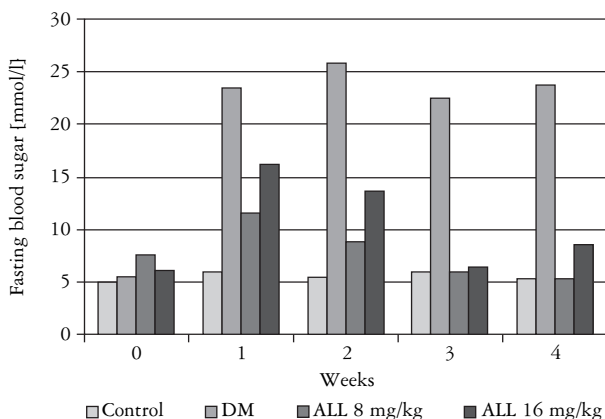


Fig. 1. Fasting blood sugar for all rat groups with/without treatment

## Results

### Immunological and biochemical findings

The diabetic animals exhibited consistent hyperglycemia (Fig. 1). Meanwhile, allicin treatment caused a decrease in the elevated serum glucose (Fig. 1), and however, Fig. 2 demonstrated the changes to the body weight of all rat groups during the experiment. Rat body weight was elevated by treatment in both allicin doses.

There is an increase (P = 0.001) in the lowered serum insulin concentrations in STZ-induced diabetic rats by the end of the experiment (Fig. 3).

After induction of IDDM, the diabetic animals showed an increase in the levels of anti-islet cell auto antibodies ICA (Fig. 4), however, by the end of the experiment the allicin treatment specially in high doses diabetic group significantly (P = 0.001) decreased the elevated ICA levels (Fig. 4).

### Histological findings

The histological findings in the histological sections of pancreatic tissues stained with H&E in the normal control rats (GA) showed normal structure (Fig. 5A). The islets of Langerhans appeared regular in shape surrounded by a thin capsule of connective tissue with lightly stained round clusters of cells embedded in the exocrine tissue. While in STZ diabetic rats with no treatment (GB) (Fig. 5B), the findings were degenerative and necrotic changes, and shrinkage in the islets of Langerhans. The islets were relatively small, atrophied, and showed a reduction in the number of polygonal islet cells. The nucleus of necrotic cells indicated either pyknosis or marginal hyperchromasia. There was mostly hydropic degeneration and degranulation in the cytoplasm of the degenerative and necrotic cells, while some of the cells with a pyknotic nucleus had a dark eosinophilic cytoplasm.

Meanwhile, islets of Langerhans in low dose allicin treated rats (8 mg/kg) (GC) (Fig. 5C) revealed light-

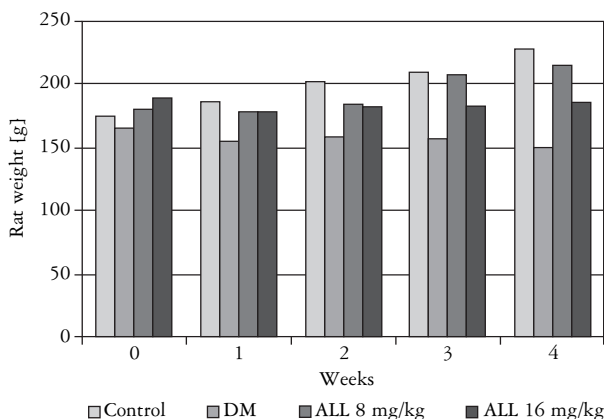
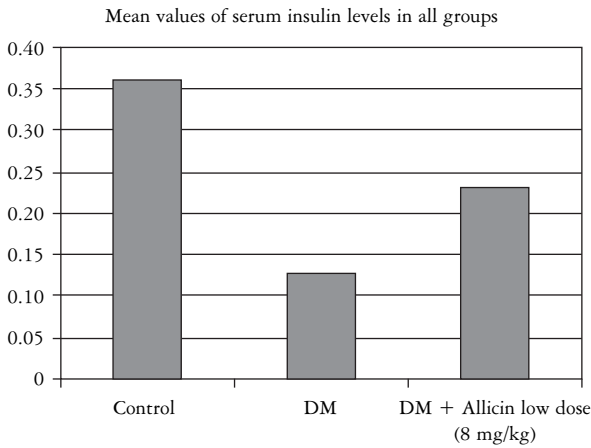
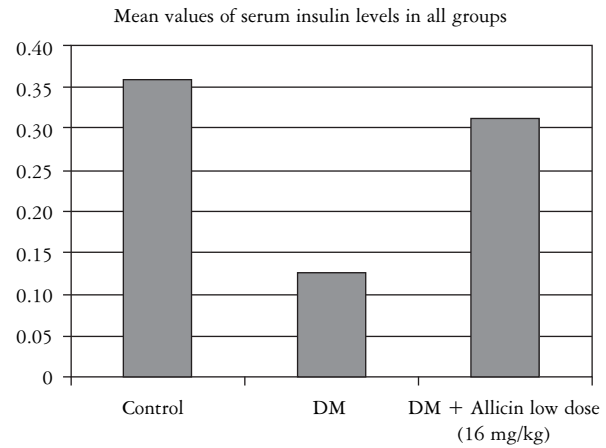


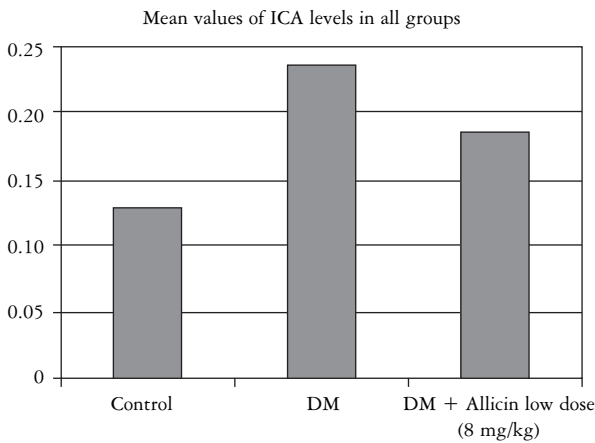
Fig. 2. Body weight for all rat groups with/without treatment



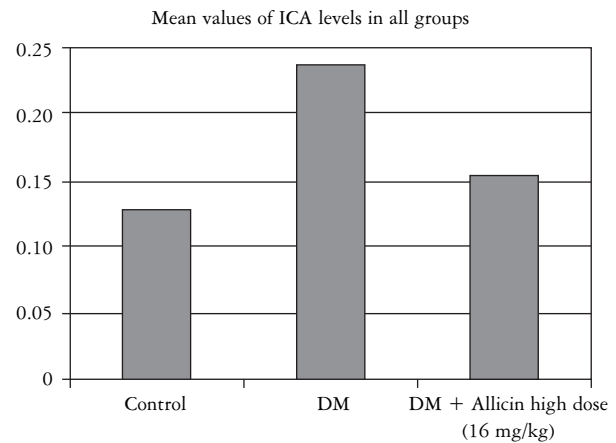
**Fig. 3A.** Effect of allicin (low dose 8 mg/kg) administration on serum insulin levels



**Fig. 3B.** Effect of allicin (high dose 16 mg/kg) administration on serum insulin levels



**Fig. 4A.** Effect of allicin (low dose 8 mg/kg) administration on the ICA levels



**Fig. 4B.** Effect of allicin (high dose 16 mg/kg) administration on the ICA levels

ly stained, elongated islets of similar size to those seen in the control group (GA) but still some islets showed disrupted cytoplasm in certain areas displaying small vacuoles, infiltration of the islet by the exocrine acinar. This figure will be much better with a high dose of allicin in the last group (GD) (Fig. 5D) since there was protection to the majority of the Langerhans islet cells which appeared regular in shape and so much similar to the normal shape in GA.

## Discussion

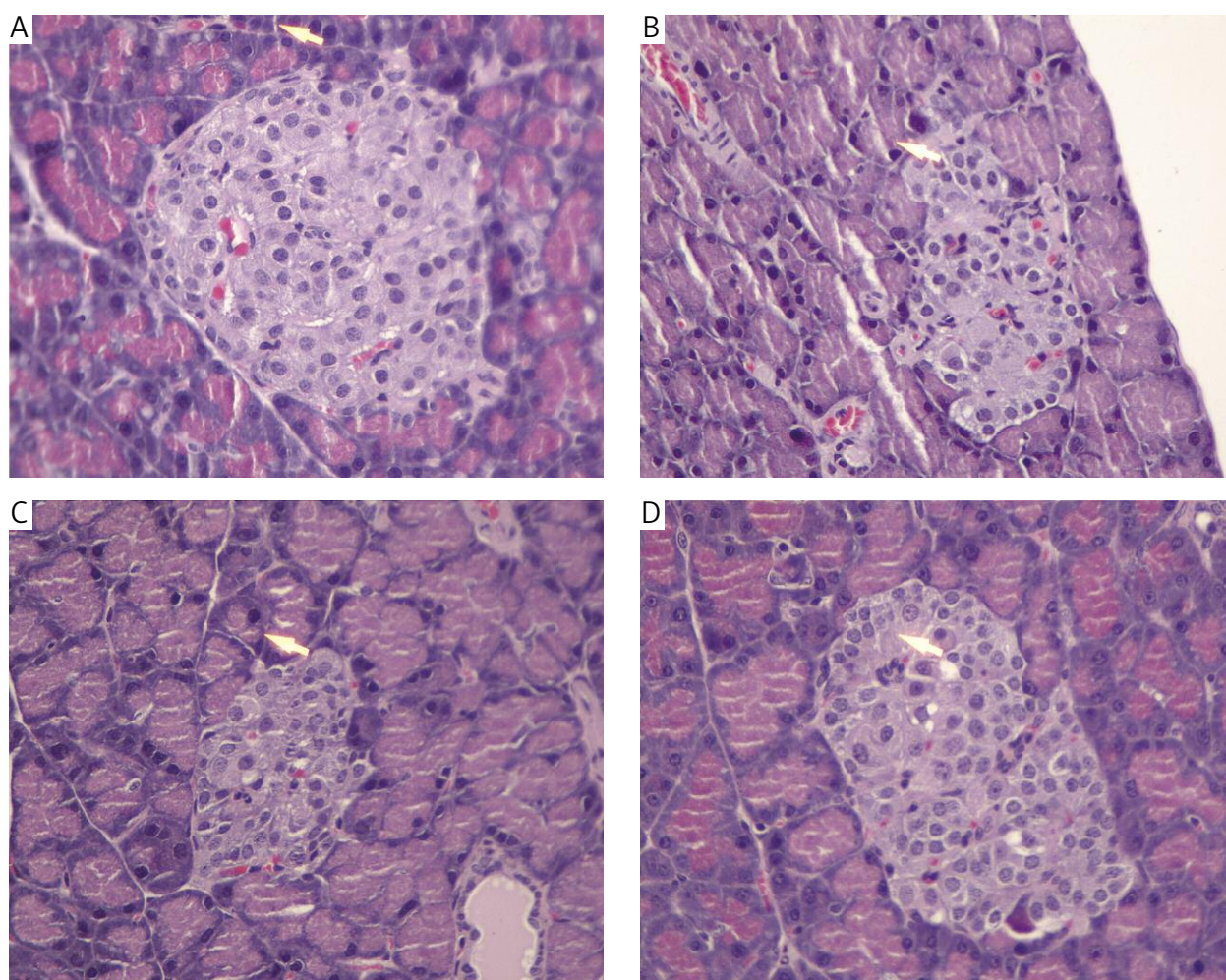
This study demonstrates for the first time, to our knowledge, the effect of allicin, the active component of garlic on the islet cell autoantibodies (ICA) production in IDDM with additional biochemical and histological evidence.

Our principal findings are: 1) daily intraperitoneal administration of allicin (either low dose of 8 mg/kg or high dose of 16 mg/kg) for up to 30 days to type 1 diabetic rats effectively reduces levels of anti-islet cell antibod-

ies which are the main antibodies produced in the autoimmune process of the disease; 2) the elevated hyperglycemia was reduced under the effect of allicin; and 3) reduced level of insulin due to damaged Langerhans islet cell was significantly increased in the serum due to the repairing tissue process due to allicin administration.

While the therapeutic anti-diabetic effects of the whole garlic or allicin alone on type 2 diabetes mellitus are numerous and well-documented, evidence presented in this study actually shows a novel immunomodulatory effect of allicin in IDDM of STZ-induced diabetic rats. This finding is made more interesting by the fact that many studies [17-20] have shown that allicin is hypoglycemic in both types of diabetes through biochemical evidence while in this study we have proved that allicin has a significant effect on the production of main autoimmune antibodies in IDDM that lead to affect and stop the disease autoimmune process of IDDM.

Studies have determined that the consequence of ICA autoantibodies in IDDM is a destruction



**Fig. 5.** Microphotographs of pancreatic tissue. HE 420. A – control group, showing normal cells in the islets of Langerhans; B – diabetic group. Shrunken islets of Langerhans displaying degenerative and necrotic changes in diabetic rats with no treatment; C – allicin-treated group (8 mg/kg): Allicin protected the majority of cells in the islets of Langerhans; D – allicin-treated group 16 mg/kg; looks like a normal control group)

of the insulin-producing  $\beta$  cells of the islets of Langerhans' cells and an absence or deficiency of circulating insulin [26]. The autoimmune attack of these antibodies appears to destroy  $\beta$  cells selectively [1, 26]. Researchers have considered that ICA serum autoantibodies are an important hallmark of this disease, and assays for these islet cell antibodies have facilitated the investigation and understanding of several facets in the pathogenesis of autoimmune diabetes. Their applications have begun to extend into clinical practice and have opened new avenues for early preclinical prediction and preventive prophylaxis in IDDM [5-26].

The immunomodulatory and immunostimulatory effects of allicin alone or within garlic were reported in many studies. It potentially induces lymphocyte proliferation and macrophage phagocytosis, stimulates the infiltration of macrophages and lymphocytes in transplanted tumors, induces splenic hypertrophy, stimulates release of interleukin-2, tumor necrosis factor  $\alpha$  and interferon- $\gamma$  and enhances natural killer cell and

lymphokine-activated killer cell activity. These activities reflect effective stimulation of the immune response [27, 28]. Meanwhile, because certain diseases can be caused by immune dysfunction, modification of immune functions by garlic may contribute to the treatment and prevention of diseases. Thus, some pharmacologic effects of garlic might be mediated through immunomodification [27-30].

The present study showed additional biochemical evidence apart from that of the immunological effect of allicin. This result indicates that allicin affects body weight, blood glucose and insulin level. However, most of the studies showed that garlic can reduce blood glucose levels in diabetic mice, rats and rabbits [31-34]. It is not clear how garlic and allicin actually work in alleviating hyperglycemia. The hypoglycemic action of allicin could possibly be due to an increase in pancreatic secretion of insulin from  $\beta$ -cells, release of bound insulin or enhancement of insulin sensitivity. It has been previously suggested that allicin can enhance serum in-

sulin by effectively combining with compounds like cysteine [35].

In the current study we examined the effects of alliin on cell damage in IDDM in STZ-induced rats in concordance with immunological and biochemical effects. STZ is cytotoxic to  $\beta$ -cells [36]. Although the  $\beta$ -cell cytotoxic action of STZ is not fully understood, it is thought to be mediated by the inhibition of free radical scavenger-enzymes, which enhances the production of superoxide radicals [36]. In the present study, almost all of the insulin-producing  $\beta$ -cells were degranulated, degenerated, or necrosed in the STZ-treated rats (Fig. 3B), which led to a decrease in insulin secretion and an increase in blood glucose levels. Streptozotocin induced a significant decrease in the area of insulin immunoreactive  $\beta$ -cells. Streptozotocin causes IDDM. In our study, alliin treatment protected the majority of the Langerhans islet cells. Nevertheless, in the low doses (8 mg/kg) we observed light hydropic degeneration, degranulation, and necrosis in some cells (Fig. 5C) but with high dose (16 mg/kg), alliin prevented degeneration of  $\beta$ -cells and a picture like normal appeared (Fig. 5D). Alliin treatment increased the area of insulin immunoreactive  $\beta$ -cells significantly.

The goals of treatment of autoimmune diseases are to reduce symptoms, control the autoimmune process and maintain the body's ability to fight disease [37]. Some patients may need supplements to replace a hormone (like insulin injections in IDDM [38] or immunosuppressive medicines include corticosteroids to control or reduce the immune system's response, but these medicines often cause a lot of side effects [37]. Our findings suggest that alliin treatment has a therapeutic protective effect against autoimmune reactions in IDDM and immune defense in IDDM can be significantly improved by the administration of alliin. The data may provide new strategies for using alliin to be recommended in the clinical management, control, and prevention of IDDM.

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*The authors declare no conflict of interest.*

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