ORIGINAL PAPER

BREAST CARCINOMA GRADING ON CORE NEEDLE BIOPSY - TO GRADE OR NOT TO GRADE?

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Core needle biopsy (CNB) is well established as an important diagnostic tool in diagnosing breast cancer and it is now considered the initial method of choice for diagnosing breast disease and the basis for the treatment planning. The concordance rate between CNB and surgical excision specimen in determination of histological grade (HG) varies widely across literature, ranging from 59-91%. The aim of our study was to investigate the level of concordance between CNB and surgical excision specimen for the determination of HG for breast cancer patients. The study population included 157 women with a breast tumor who underwent a core needle biopsy for breast carcinoma and a subsequent surgical excision of the tumor. The concordance level between core needle biopsy and surgical resection specimen for overall histologic grading was 73%: for tubule formation – 71%, for nuclear pleomorphism – 91%, for the mitotic index – 59%. Our study shows that our institution's histologic grading of CNBs and surgical excisions shows a fairly good correlation and is useful for the planning of treatment.

Key words: breast cancer, grading, core needle biopsy.

Introduction

Breast cancer is the most common cancer affecting women worldwide [1, 2]. It is also the leading cause of cancer deaths among women [3]. Although the incidence of breast cancer has been increasing since the implementation of mass mammography screening and continues to grow due to the aging population, mortality has decreased over the past years [4]. This decrease in mortality rate is in part due to the earlier detection of cancer with mammography screening, as well as the development of more effective treatments [5, 6]. Presently non-operative diagnosis of breast lesions comprises "triple assessment" based on physical examination, imaging (mammography and/or ultrasound), and pathology [7]. Since 2015, the European Society of Medical Oncology guidelines for the clinical practice of breast cancer require that patients suspicious for malignancy have a pathological diagnosis performed by core-needle biopsy (CNB) before starting any treatment [8]. This allows for the personalization of the approach to the oncological patient and the determination of the basic prognostic and predictive factors necessary to make the right therapeutic decision and start surgical treatment or systemic therapy.

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In contrast to fine needle aspiration (FNA), CNB provides architectural information allowing for the evaluation of prognostic and predictive factors for breast cancer, including histological grade (HG) – one of three prognostic factors used to calculate the Nottingham Prognostic Index [9-13]. The advantages of CNB over FNA include also a more definitive histological diagnosis, differentiation between in situ and invasive tumors, and the possibility of molecular profiling and assessment of biomarkers [14].

In comparison to an excisional biopsy (EB), a core needle biopsy is a simple, cost effective, and less invasive procedure with a low complication rate that has been proven to have a high accuracy, sensitivity, and specificity in diagnosing breast cancer [15]. Previous studies have shown CNB to be almost as accurate as open excisional biopsy in diagnosing breast disease [16], and to have an excellent agreement between diagnosis made by examination of CNB and surgical specimens [17, 18]. This concordance is also observed in the context of histopathological biomarkers such as ER, PR and HER2, indicating that retesting for surgical excision may not be necessary [19, 20]. All this means that CNB is well established as an important diagnostic tool in diagnosing breast cancer and it is now considered the initial method of choice for diagnosing breast disease [15, 16, 18, 21-24]. In many cases, CNB may be the only cancer sample for patients showing complete pathological response to neoadjuvant therapy, which is increasingly used to down-stage primary tumors prior to breast-conserving therapy, and to reduce the risk of metastasis [25].

One of the powerful independent prognostic and predictive factor assessed in the breast cancer tissue is the histological grade. Several studies have previously described the concordance rate between CNB and EB specimen in determination of HG [26-29]. Many studies has also been conducted to find out the cytological grading system that correlates well with histological grading. However, the concordance rate previously ascribed to overall grade is controversial and varies widely across literature, ranging from 59-91% [26, 30-33]. This may potentially exclude some patients that would benefit from neoadjuvant therapy. This fact makes the determination of the actual concordance of HG in the core biopsy sample and the excisional biopsy specimen an important clinical problem.

To the best of our knowledge, no study investigating the level of agreement between CNB and surgical excision specimen for the determination of HG for breast cancer patients has been performed in Poland. The aim of our study was to evaluate the relevant data recorded at our institution and compare our results to those described in previous literature.

Material and methods

The study population included 157 women with a breast cancer who underwent a core needle biopsy and a subsequent surgical excision of the tumor. Samples were routinely processed and embedded in paraffin wax (FFPE). All specimens were assessed in 2 µm HE-stained FFPE sections as part of the routine reporting, by experienced breast pathologists. HG was assessed only in core needle biopsies containing at least 10 well preserved HPF (×400, field diameter 0.55 mm) with invasive tumor. The total score of HG consisted of the evaluation of individual histological features: tubule formation, nuclear pleomorphism, and mitotic count. Evaluation of tubule formation was based on the percentage of cells in the tumor that have tube-shaped structures with clear central lumina: score 1 - over 75% of the cancer was composed of tubular structures, score 2-10-75% of the tumor had a tubular pattern and score 3-less than 10% of the tumor contained tubules. The assessment of nuclear pleomorphism in areas with the greatest atypia was based on a qualitative analysis of the nuclear morphology of the tumor, assessed microscopically on a scale of 1 to 3, reflecting increasing differences in appearance compared to normal epithelium:

- nuclear pleomorphism score 1 nuclei similar in size to nuclei of normal epithelial cells ($< 1.5 \times$ the size of normal epithelial cell nuclei), minimal nuclear variation in size and shape, small regular uniform cells, invisible or very small nucleoli,
- nuclear pleomorphism score 2 nuclei larger (1,5-2 × the size of normal epithelial cell nuclei), moderate nuclear variation in size and shape, visible but small nucleoli,
- nuclear pleomorphism score 3 nuclei larger (> 2 × the size of normal epithelial cell nuclei), marked nuclear variation in size and shape, large nucleoli.

The mitotic counts were divided into three mitotic scores: mitotic score 1 for 0-8 mitoses/10 HPF, mitotic score 2 for 9-17 mitoses/10 HPF and mitotic score 3 for \geq 18 mitoses/10 HPF (field diameter 0,55 mm).

Both materials were evaluated for the determination of histological grade depending on the sum of the points obtained: G1 (3-5 points), G2 (6-7 points) and G3 (8-9 points).

Samples from patients before and after neoadjuvant chemo- or hormone therapy were excluded from the study.

The illustrations were done from the whole slide images using Medlan scan viewer.

The degree of concordance between CNB and surgical excision specimen for the determination of tumor grade was assessed by Cohen's κ coefficient. The κ coefficient used was $\kappa = 0.703, 95\%$

INTERPRETATION
No concordance
Very poor concordance
Good concordance
Very good concordance
100% concordance

Table I. General criteria for assessing the degree of concordance based on $\kappa\mbox{-}value$

 Table II. Concordance between CNB and surgical excision

 for histological grade

		SURGICAL EXCISION			
		G 1	G2	G3	TOTAL
CNI	3				
(G1	21	12	1	34
(G2	5	52	20	77
(G3	1	3	40	44
,	Fotal	27	67	61	155

CI: 0.5728-0.8332. Table I describes the general criteria used to assess the degree of concordance based on κ -value [34-36]. A weighted coefficient was applied in the analysis in order to assign a particular weight for each stage. The higher the stage, the higher the weight (ordinal scale: the higher the level, the worse condition) (Table I).

Results

Retrospective comparison of medical records and pathological reports revealed that CNB correctly predicted the histological grade in 113 of 155 cases (73%). The level of agreement between core needle biopsy and surgical resection specimen for overall histologic grading was 73% (113 of 155 cases). CNB correctly predicted the grade of the surgical excision specimen in 21 cases for grade 1 tumors ($\kappa = 0.525$, 95% CI: 0.36340-0.6818 F, 52 cases for grade 2 tumors ($\kappa = 0.5652$, 95% CI: 0.458-0.667), and 40 cases for grade 3 tumors ($\kappa = 0.6154$, 95% CI: 0.4862-0.7309). The highest level of agreement was observed in grade 3 malignancies (Table II).

The concordance rate for tubule formation was 71% (126 out of 155 cases) with a κ of 0.7489, 95% CI: 0.5868-0.911. Analyzing tubule formation scores separately, the concordance rates were 100% (8 of 8 cases) for grade 1 tumors, 72.4% (42 of 58) for grade 2 tumors, and 85.4% (76 of 89) for grade 3 tumors. The highest concordance rate for tubule formation

Table III. Concordance between CNB and surgical excision for tubule formation

		SURGICAL EXCISION			
		G 1	G2	G3	TOTAL
CNB					
	G1	8	0	0	8
(G2	3	42	13	58
	G3	0	13	76	89
	Total	11	55	89	155

Table IV. Concordance between CNB and surgical excision for nuclear pleomorphism

		SURGICAL EXCISION			
		G 1	G2	G3	TOTAL
CN	1B				
	G1	1	1	0	2
-	G2	0	100	12	112
-	G3	0	1	40	41
	Total	1	102	52	155

Table V. Concordance between CNB and surgical excision for mitotic index

		SURGICAL EXCISION			
		G 1	G2	G3	TOTAL
CN	JB				
_	G1	30	24	15	69
	G2	7	30	13	50
-	G3	0	4	32	36
	Total	37	58	60	155

Table VI. Comparison of tumor grade between CNB and surgical excision

	Grade (%) ($N = 155$)
CNB = surgical excision	113 (73%)
Surgical excision > CNB	33 (21%)
CNB > surgical excision	9 (6%)

as an individual parameter of HG was observed amongst tumors assigned a score of 1 (Table III).

The score for nuclear pleomorphism was concordant in 91% of all cases (141 of 155, $\kappa = 0.801$, 95% CI: 0.5556-1). The concordance rates for the individual grades were 50% (1 of 2 cases) for grade 1, 89.3% (100 of 112) for grade 2, and 97.6% (40 of 41)

AUTHORS	No.	GRADE (%)	TUBULE FORMATION (%)	NUCLEAR PLEOMORPHISM (%)	Mitoses (%)
Daveau et al. (2014)	350	78 grade 1	75	66.5	75
		68 grade 2			
		95 grade 3			
		75 combined grade			
Lorgis et al. (2011)	175	75.4			
Ough et al. (2011)	209	63			61
Park <i>et al</i> . (2009)	104	80.8			
Ozdemir et al. (2007)	201	77.8 grade 1			
		69.2 grade 2			
		61.5 grade 3			
		68.8 combined grade			
Usami et al. (2007)	111	75		61	
Cahill et al. (2006)	95	77			
Burge et al. (2006)	87	81 grade 1			
		83 grade 2			
		65 grade 3			
		77 combined grade			
Badoual et al. (2005)	110	73.1	78.5	79.6	60.2
Usami et al. (2005)	22	80		54	
Monticciolo (2005)	288	74.3		76.6	
Deshpande et al. (2005)	105	100 grade 1			
		71 grade 2			
		50 grade 3			
		75 combined grade			
O'Leary et al. (2004)	113	61.6	55.6	57.4	59.4
Andrade and Gobbi (2004)	120	59	54.7	58.9	62.1
Harris et al. (2003)	500	60 grade 1	82	73	58
		60 grade 2			
		84 grade 3			
		67 combined grade			
Connor et al. (2002)	44	64			
McIntosh et al. (2002)	133	91			
Shannon <i>et al</i> . (2001)	734	75 grade 1			
		70 grade 2			
		86 grade 3			
Sharifi et al. (1999)	79	75			

Table VII. Concordance rate (%) between core biopsies and subsequent surgical excisions in the literature [46]

for grade 3. The highest level of agreement for nuclear pleomorphism was observed in tumors assigned a score of 3 (Table IV).

The mitotic index was concordant in 59% of all cases (92 of 155, $\kappa = 0.4901$, 95% CI: 0.363-0.6172). Only 30 of 69 cases were correctly deter-

mined to be grade 1 for mitotic index in both core biopsy and surgical excision specimens (concordance rate 43.5%). The concordance levels for grade 2 and grade 3 mitotic indexes were 60% (30 of 50 cases) and 88.9% (32 of 36 cases), respectively. The greatest rate of concordance for mitotic index was in tumors

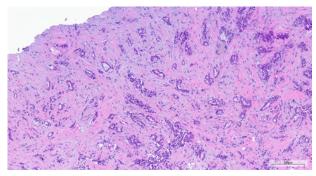


Fig. 1. HG 1 on CNB and HG 1 on ES in the same patient (HE stain, magnification $100 \times$)

assigned a score of 3 (Table V). Comparison of HG based on core needle biopsy and excisional biopsy is shown in Table VI, VII. Figures 1-3 demonstrate comparison of tumor HG on CNB and EB.

Discussion

Accurate evaluation of breast cancer on biopsy samples is of crucial importance to guide therapeutic decisions. This fact justifies a thorough assessment of the concordance between the prognostic and predictive factors routinely determined in the biopsy material and in the excised tumor. One of the factors considered in this assessment is histological grade.

HG in breast carcinoma is the combination of three histological features: tubule formation, nuclear pleomorphism, and mitotic count. Since 1991, histological grade as part of the Nottingham (Elston Ellis) modification of the Scarff-Bloom-Richardson grading system, also known as the Nottingham Grading System (NGS), has continuously proven to be a powerful prognostic factor in guiding the management of breast cancer patients [26, 36-40]. In comparison to novel prognostic molecular tests, histological tumor grade remains an easily accessible and highly accurate alternative method for assessing tumor morphology and biological characteristics, as well as patient prognosis.

In breast cancer, histological evaluation performed on tumor samples after breast-conserving surgery or mastectomy is the standard of care [41, 42]. However, HG in excision specimens may change or become difficult if not impossible to assess following neoadjuvant chemotherapy. In the case of a complete pathological response, as in metastatic patients who are not amenable to surgical resection, core needle biopsy specimen is the only available sample of primary tumor. In such circumstances, it becomes essential to know the HG initial result to determine the prognosis of the patient. Similarly, it may be necessary to know the grade of breast

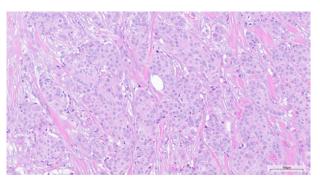


Fig. 2. HG 2 on CNB and HG 1 on ES in the same patient (HE stain, magnification $200 \times$)

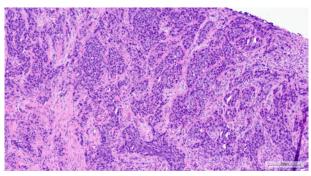


Fig. 3. HG 3 on CNB and HG 3 on ES in the same patient (HE stain, magnification $100 \times$)

carcinoma before tumor excision, to decide about neoadjuvant treatment [43]. Optimally, the result obtained from the CNB should be consistent with the surgical sample. HG evaluated in core needle specimens, although widely used, has shown a variable concordance with the final histological grade in previous studies [31-33, 44, 45].

In presented study we evaluated the concordance rate of breast carcinoma histological grade and individual parameters of HG on CNB and subsequent surgical specimen. Our results showed that histological grade of CNB accurately predicted that of surgical specimen in 113 of 155 cases (73%) with a $\kappa\text{-value}$ of 0.703. Earlier studies have obtained similar results ranging from 51-91% [31-33, 44, 45], with pooled agreement 71.1% calculated in meta-analysis [46]. Among 33 of 42 (79%) discordant cases, the grade was higher in the surgical excision than in the CNB. This accounted for 21% of the full 27% of discordant cases in our study. In 9 of 42 (21%) discordant cases, the grade was higher in the CNB than in the surgical excision. This composed 6% of the overall discordance. These results corresponds to the noted in the literature, showing that underestimation occurs more frequently than overestimation [46]. There are various explanations for discordance between CNB and HG profiles in breast cancer, including tumor heterogeneity and pre-analytic variation [20].

Analysis of the separate histological grades revealed concordance rates of 62% (21 of 34) in grade 1, 68% (52 of 77) in grade 2, and 91% (40 of 44) in grade 3 tumors. These results are similar to those reported by Daveau et al., Harris et al., Shannon et al., and Focke et al. whose values for grade 1, grade 2 and grade 3 tumors vary from 60-78%, 60-70%, to 84-99%, respectively [30, 41, 47, 48]. The higher concordance rates we observed in high grade tumors (grade 3; 91%) in our study is consistent with previous reports. However, the opposite results have been found in other studies, notably Desphpande et al., Burge et al., and Ozdemir et al. For those studies, the concordance rates for grade 1, grade 2, and grade 3 tumors were 77.8-100%, 69.2-83, and 50-65%, respectively [33, 49, 50].

We also assessed three morphological features that constitute the grade for an individual tumor. Regarding tubule formation, nuclear pleomorphism, and mitotic index, the concordance rate was 71% ($\kappa =$ 0.7489), 91% ($\kappa = 0.801$), and 59% ($\kappa = 0.4901$), respectively. Previous studies have shown similar concordance rates ranging from 54.7-82%, 54-79.6%, and 58-75% for tubule formation, nuclear pleomorphism, & mitotic indices, respectively [26, 30-33, 51]. The greatest discrepancy between concordance rates for individual parameters of HG in our study was observed in the mitotic index. A possible reason for this could be the inevitability of sampling error. In comparison to a surgical excision specimen, a CNB offers much less tissue for histological evaluation and may not guarantee an adequate specimen from the periphery of the tumor where the active growth would likely contain the most mitotic activity [37].

The majority of obtained results fall within the ranges of concordance rates reported in earlier studies, and in a similar fashion demonstrate that the under grading of breast carcinoma on CNB is largely due to the underestimation of mitotic counts and the overestimation of nuclear pleomorphism [37, 52, 53]. The reported in presented study and in the literature discordance in tumor grading between CNB and resection specimens from breast cancer affects the indication for adjuvant therapy in only a small minority of patients with invasive carcinoma [54].

In this study, we performed a retrospective analysis of HG concordance between CNB and breast cancer EB specimens, comparable to others reported in the literature. Our study focused on clinically relevant end points based on discordance, incluour data was collected from an academic and tertiary referral center. However, the study also has inherent limitations given its retrospective design and relatively small sample size. Improvements in concordance scores can only be expected from increasing the size and representativeness of biopsy specimens (e.g. using vacuum-assisted biopsy) and gaining more experience of the breast pathologist.

Conclusions

Presented study shows that our institution's histologic grading of CNBs and surgical excisions shows a fairly good correlation and is consistent with findings in previous reports. Despite the inevitable limitations of CNB, it is an effective method for diagnosing breast cancer and managing treatment options. Assessment of tumor grade by CNB is useful for the planning of treatment, so in authors opinion it is worthy to implement it in daily practice.

Institutional Review Board Statement: Ethical review and approval were waived for this study due to analysis based on archival data only [55].

The authors declare no conflict of interest.

References

- World Health Organization (WHO). Breast Cancer: prevention and control. Available at: http://www.who.int/cancer/ detection/breastcancer/en/index1.html (accessed on January 9, 2018).
- Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015; 136: E359-E386.
- Sung H, Ferlay J, Siegel RL, et al. Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. Cancer J Clin 2021; 71: 209-249.
- Kohler BA, Sherman RL, Howlander N, et al. Annual Report to the Nation on the Status of Cancer, 1975-2011, Featuring Incidence of Breast Cancer Subtypes by Race/Ethnicity, Poverty, and State. J Natl Cancer Inst 2015; 107: djv048.
- Independent UK Panel on Breast Cancer Screening. The benefits and harms of breast cancer screening: an independent review. Lancet 2012; 380: 1778.
- Ferlay J, Parkin DM, Steliarova-Foucher E. Estimates of cancer incidence and mortality in Europe in 2008. Eur J Cancer 2010; 46: 765-781.
- Kooistra B, Wauters C, Strobbe L, Wobbes T. Preoperative cytological and histological diagnosis of breast lesions: A critical review. Eur J Surg Oncol 2010; 36: 934-940.
- Senkus E, Kyriakides S, Ohno S, et al.; ESMO Guidelines Committee. Primary breast cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2015; 26: 8-30.
- Denley H, Pinder SE, Elston CW, et al. Preoperative assessment of prognostic factors in breast cancer. J Clin Pathol 2001; 54: 20-24.
- 10. Wang M, He X, Chang Y, et al. A sensitivity and specificity comparison of fine needle aspiration cytology and core needle biopsy in evaluation of suspicious breast lesions: A systematic review and meta-analysis. Breast 2017; 31: 157-166.
- Haybittle JL, Blamey RW, Elston CW, et al. A prognostic index in primary breast cancer. Br J Cancer 1982; 45: 361-366.
- 12. Todd JH, Dowle C, Williams MR, et al. Confirmation of a prognostic index in primary breast cancer. Br J Cancer 1987; 56: 489-492.
- 13. Blamey RW, Ellis IO, Pinder SE, et al. Survival of invasive breast cancer according to the Nottingham Prognostic In-

dex in cases diagnosed in 1990-1999. Eur J Cancer 2007; 43: 1548-1555.

- Britton PD. Fine needle aspiration or core biopsy. Breast 1999; 8: 1-4.
- 15. Pettine S, Place R, Babu S, et al. Stereotactic breast biopsy is accurate, minimally invasive, and cost effective. Am J Surg 1996; 171: 474-476.
- 16. Bruening W, Fontanarosa J, Tipton K, et al. Systematic review: comparative effectiveness of core-needle and open surgical biopsy to diagnose breast lesions. Ann Intern Med 2010; 152: 238-246.
- 17. Dahlstrom JE, Jain S, Sutton T, et al. Diagnostic accuracy of stereotactic core biopsy in a mammographic breast cancer screening programme. Histopathology 1996; 28: 421-427.
- Di Loreto C, Puglisi F, Rimondi G, et al. Large core biopsy for diagnostic and prognostic evaluation of invasive breast carcinomas. Eur J Cancer 1996; 32A: 1693-1700.
- Shanmugalingam A, Hitos K, Hegde S, et al. Concordance between core needle biopsy and surgical excision for breast cancer tumor grade and biomarkers. Breast Cancer Res Treat 2022; 193: 151-159.
- 20. Asogan AB, Hong GS, Arni Prabhakaran SK. Concordance between core needle biopsy and surgical specimen for oestrogen receptor, progesterone receptor and human epidermal growth factor receptor 2 status in breast cancer. Singapore Med J 2017; 58: 145-149.
- Pinder SE, Elston CW, Ellis IO. The role of pre-operative diagnosis in breast cancer. Histopathology 1996; 28: 563–566.
- 22. Pijnappel RM, van Dalen A, Borel Rinkes IH, et al. The diagnostic accuracy of core biopsy in palpable and non-palpable breast lesions. Eur J Radiol 1997; 24: 120-123.
- Crowe JP, Patrick RJ, Rybicki LA, et al. Does ultrasound core breast biopsy predict histologic finding on excisional biopsy? Am J Surg 2003; 186: 397-399.
- 24. Cipolla C, Fricano S, Vieni S, et al. Validity of needle core biopsy in the histological characterisation of mammary lesions. Breast 2006; 15: 76-80.
- 25. Kwok TC, Rakha EA, Lee AH, et al. Histological grading of breast cancer on needle core biopsy: the role of immunohistochemical assessment of proliferation. Histopathology 2010; 57: 212-219.
- 26. Rakha EA, El-Sayed ME, Lee AH, et al. Prognostic significance of Nottingham histologic grade in invasive breast carcinoma. J Clin Oncol 2008; 26: 3153-3158.
- 27. Amat S, Penault-Llorca F, Cure H, et al. Scarff-Bloom-Richardson (SBR) grading: a pleiotropic marker of chemosensitivity in invasive ductal breast carcinomas treated by neoadjuvant chemotherapy. Int J Oncol 2002; 20: 791-796.
- 28. Petit T, Wilt M, Velten M, et al. Comparative value of tumour grade, hormonal receptors, Ki-67, HER-2 and topoisomearase II alpha status as predictive markers in breast cancer patients treated with neoadjuvant anthracycline-based chemotherapy. Eur J Cancer 2004; 40: 205-211.
- 29. Rajendran K, Sudalaimuthu M, Ganapathy S. Cytological grading of breast carcinomas and its prognostic implications. Cureus 2022; 14: e29385.
- 30. Daveau C, Baulies S, Lalloum M, et al. Histological grade concordance between diagnostic core biopsy and corresponding surgical specimen in HR-positive/HER2-negative breast carcinoma. Br J Cancer 2014; 110: 2195-2200.
- 31. Lorgis V, Algros MP, Villanueva C, et al. Discordance in early breast cancer for tumour grade, estrogen receptor, progesteron receptors and human epidermal receptor-2 status between core needle biopsy and surgical excisional primary tumour. Breast 2011; 20: 284-287.
- 32. Ough M, Velasco J, Hieken TJ. A comparative analysis of core needle biopsy and final excision for breast cancer: histology and marker expression. Am J Surg 2011; 201: 692-694.

- 33. Ozdemir A, Voyvoda NK, Gultekin S, et al. Can core biopsy be used instead of surgical biopsy in the diagnosis and prognostic factor analysis of breast carcinoma? Clin Breast Cancer 2007; 7: 791-795.
- 34. Gwet KL. Handbook of Inter-Rater Reliability: The Definitive Guide to Measuring the Extent of Agreement Among Multiple Raters. 3rd ed. Gaithersburg, MD: Advanced Analytics, LLC; 2012.
- 35. Measurement Systems Analysis: Reference Manual. 4th ed. AIAG-Work Group, Daimler Chrysler Corporation, Ford Motor Company, General Motors Corporation, 2010.
- 36. Hamrol A. Zarządzanie jakością z przykładami. Pub. PWN, Warszawa, 2008.
- 37. Elston CW, Ellis IO. Pathological prognostic factors in breast cancer. I. The value of histological grade in breast cancer: experience from a large study with long-term follow-up. Histopathology 1991; 19: 403-410.
- Elston CW, Ellis IO, Pinder SE. Pathological prognostic factors in breast cancer. Crit Rev Oncol Hematol 1999; 31: 209-223.
- 39. Anderson TJ, Alexander FE, Lamb J, et al. Pathology characteristics that optimize outcome prediction of a breast screening trial. Br J Cancer 2000; 83: 487-492.
- Frkovic-Grazio S, Bracko M. Long term prognostic value of Nottingham histological grade and its components in early (pT1N0M0) breast carcinoma. J Clin Pathol 2002; 55: 88-92.
- 41. Focke CM, Decker T, van Diest J. The reliability of histological grade in breast cancer core needle biopsies depends on biopsy size: a comparative study with subsequent surgical excisions. Histopathology 2016; 69: 1047-1054.
- 42. Rakha EA, Reis-Filho JS, Baehner F, et al. Breast cancer prognostic classification in the molecular era: the role of histological grade. Breast Cancer Res 2010, 12: 207.
- 43. Reinert T, de Souza AB, Sartori GP, et al. Highlights of the 17th St Gallen International Breast Cancer Conference 2021: customising local and systemic therapies. Ecancermedicalscience 2021; 15: 1236.
- 44. Andrade VP, Gobbi H. Accuracy of typing and grading invasive mammary carcinomas on core needle biopsy compared with the excisional specimen. Virchows Arch 2004; 445: 597-602.
- 45. O'Shea AM, Rakha EA, Hodi Z, et al. Histological grade of invasive carcinoma of the breast assessed on needle core biopsy – modifications to mitotic count assessment to improve agreement with surgical specimens. Histopathology 2011; 59: 543-548.
- 46. Knuttel FM, Menezes GLG, van Diest PJ, et al. Meta-analysis of the concordance of histological grade of breast cancer between core needle biopsy and surgical excision specimen. Br J Surg 2016; 103: 644-655.
- 47. Harris GC, Denley HE, Pinder SE, et al. Correlation of histologic prognostic factors in core biopsies and therapeutic excisions of invasive breast carcinoma. Am J Surg Pathol 2003; 27: 11-15.
- 48. Shannon J, Douglas-Jones AG, Dallimore NS. Conversion to core biopsy in preoperative diagnosis of breast lesions: is it justified by results? J Clin Pathol 2001; 54: 762-765.
- 49. Deshpande A, Garud T, Holt SD. Core biopsy as a tool in planning the management of invasive breast cancer. World J Surg Oncol 2005; 3: 1.
- 50. Burge CN, Chang HR, Apple SK. Do the histologic features and results of breast cancer biomarker studies differ between core biopsy and surgical excision specimens? Breast 2006; 15: 167-172.
- 51. Park SY, Kim KS, Lee TG, et al. The accuracy of preoperative core biopsy in determining histologic grade, hormone receptors, and human epidermal growth factor receptor 2 status in invasive breast cancer. Am J Surg 2009; 197: 266-269.

- 52. Thunnissen FB, Ambergen AW, Koss M, et al. Mitotic counting in surgical pathology: sampling bias, heterogeneity and statistical uncertainty. Histopathology 2001; 39: 1-8.
- 53. Sharifi S, Peterson MK, Baum JK, et al. Assessment of pathologic prognostic factors in breast core needle biopsies. Mod Pathol 1999; 12: 941-945.
- 54. Waaijer L, Willems SM, Verkooijen HM, et al. Impact of preoperative evaluation of tumour grade by core needle biopsy on clinical risk assessment and patient selection for adjuvant systemic treatment in breast cancer. Br J Surg 2015; 102: 1048-1055.
- 55. Marszałek A, Kubicka A, Jagiełło I, Malicka-Durczak A. Potential impact of HercepTest[™] mAb PharmDx (Dako Omnis) (ge001) in breast cancer diagnosis. Pol J Pathol 2023; 74: 82-88.

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