

Rapid diagnostic tests to detect asymptomatic malaria in primary health care facilities in hypoendemic areas

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A – Study Design, **B** – Data Collection, **C** – Statistical Analysis, **D** – Data Interpretation, **E** – Manuscript Preparation, **F** – Literature Search, **G** – Funds Collection

Summary Background. Early detection and appropriate treatment of malaria, as well as elimination of vectors, will prevent the transmission of the disease. Nowadays, malaria treatment is only administered after the illness has been laboratory confirmed using, e.g., rapid diagnostic tests (RDTs), when microscopy is not available. RDTs are commonly used in patients reporting to primary health care (PHC) providers with a fever. If left untreated, asymptomatic malaria could be a source of disease transmission and may potentially lead to an outbreak. Yet, the detection of asymptomatic malaria is difficult, as parasitological tests are only performed in symptomatic patients, i.e. those presenting with a fever.

Objectives. The aim of the study was to assess the rates of asymptomatic malaria cases in hypoendemic areas. Malaria was detected through RDTs that were performed at PHC facilities in the Batubara District in the North Sumatera Province in Indonesia.

Material and methods. Data was collected from two PHC providers and four villages with the highest rate of malaria cases. Malaria was diagnosed based on RDTs and then confirmed by microscopic examination, which remains the gold standard.

Results. The study group included 504 patients. The overall sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) of the RDTs for the diagnosis of any malarial species were 77.9%, 41.9%, 59.4% and 63.5%, respectively. The proportion of asymptomatic malaria was 22.1%. Asymptomatic malaria cases were mainly due to *Plasmodium vivax*.

Conclusions. Periodic mass blood surveys are necessary to identify asymptomatic malaria cases in hypoendemic areas.

Key words: fever, RDTs, asymptomatic malaria, PHC.

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Background

Even today, malaria is still the most common mosquito-borne disease in the Batubara District in the North Sumatera Province in Indonesia. Fluctuations of the prevalence of malaria occur frequently in many places in Indonesia [1]. The cause of fluctuations in the case of malaria is failed detection of asymptomatic malaria. Patients of asymptomatic malaria generally do not come to health care facilities. These individuals can be a source of new transmission in subsequent malaria infection and form a reservoir of parasites in vector [2]. The transmission will form the reservoir in a vector, especially in hypoendemic malaria, and could potentially be the cause of an increase in cases of malaria [3].

There have been continuous efforts to eliminate malaria in the region, yet, because cases of asymptomatic malaria are not usually treated, these efforts have failed. Asymptomatic carriage of malaria pathogens may facilitate the transmission of the disease and potentially be the cause of an outbreak and, as a consequence, a rise in the prevalence of malaria. Microscopy continues to be the 'gold standard' for malaria diagnosis; unfortunately, it is not always available at primary health care (PHC) facilities in resource-limited settings, due to its high cost, lack of skilled manpower, ac-

cessories and the reagents required [4]. Meanwhile, rapid diagnostic tests (RDTs) are being used as a diagnostic tool for malaria as a substitute for microscopic examination [5, 6].

The efficacy of RDTs is considered similar to microscopic examination in hyperendemic areas according to some experts. In addition, RDTs have been shown to be cost effective and less time consuming in the management of malaria [4]. Detection of asymptomatic malaria is an important task which is mainly carried out in PHC facilities and is the key element of the malaria elimination program.

Objectives

The aim of this study was to assess the rates of asymptomatic malaria cases in hypoendemic areas. Malaria was detected through RDTs that were performed at PHC facilities in the Batubara District in the North Sumatera Province in Indonesia.

Material and methods

The study was carried out to detect asymptomatic infections of malaria using RDTs. The data was collected from two



PHC facilities and four villages from November to December 2015. This study involved 504 patients (49.8% females and 50.2% males), from 1 to 70 years of age, living in the Batubara District. All randomly selected participants were screened using active case detection (ACD) and passive case detection (PCD). Axillary temperature was measured in all patients upon recruitment. Fever was defined as an axillary temperature $\geq 37.5^{\circ}\text{C}$. Every patient willing to participate in this study was examined using RDTs. Malaria diagnosis was confirmed by microscopic examination, which continues to be the gold standard for the detection of malaria. The RDTs used in this study included: Monotes Test Device, Malaria Pf/Pv Ag Rapid Test (whole blood), with relative sensitivity of 100%, relative specificity of 98.7% and an overall agreement of 98.9%. The sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) of the RDTs were assessed for malaria detection.

Ethics approval and authorization were obtained from the ethical committee of the Medical Faculty of the University of Sumatera Utara. Before collecting the data, each participant was clearly informed about the objective of the study, and verbal permission from the head of each household was obtained.

Results

A total of 504 patients were examined throughout the study period. There were 345 patients with fever and 159 patients without fever. The RDTs that were performed on 159 patients without fever demonstrated that 58 patients (36.5%) had asymptomatic malaria (Table 1). In the group of patients without fever, the majority was asymptomatic or presented with weakness (Table 2). Almost all cases of asymptomatic malaria were due to *Plasmodium vivax*.

Table 1. Classification of malaria

Age (years)	Sex		Fever (+)		Fever (-)	
	M	F	clinical malaria	non malaria	asymptomatic malaria	non malaria
≤ 5	21	35	18	20	4	14
6–12	63	56	51	40	12	19
13–19	57	76	52	42	25	15
20–50	98	76	77	35	14	42
> 50	14	8	7	3	3	11

Table 2. Classification of asymptomatic malaria

Symptoms	<i>P. falciparum</i> (%)	<i>P. vivax</i> (%)	<i>P. falciparum</i> + <i>P. vivax</i> (%)
Cough	0	1.7	5.2
Weakness	0	22.4	12.1
Diarrhea	0	5.2	0
Vomiting	0	8.6	0
Headache	1.7	3.4	1.7
Myalgia	0	1.7	1.7
Symptomless	1.7	22.4	10.3

The prevalence of asymptomatic malaria, based on the results of RDTs, was 22.1%. The RDTs were positive in 205/263 (77.9%; sensitivity) cases of fever and negative in 101/241 (41.9%; specificity) patients without fever. The PPV for malaria infection in patients with fever was 205/345 (59.4%), and NPV was 101/159 (63.5%). The sensitivity, specificity, PPV and NPV results of RDTs for the detection of malaria infection are presented according to age groups (Table 3).

Table 3. Accuracy of Rapid Diagnostic Test

Age (years)	Asymptomatic malaria (%)	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)
≤ 5	1.5	81.8	41.2	47.4	77.8
6–12	4.6	81	32.2	56	61.3
13–19	9.5	67.5	26.3	55.3	37.5
20–50	5.3	84.6	54.5	68.8	75
> 50	1.1	70	78.6	70	78.6

Discussion

Many countries have increased efforts to control malaria by introducing elimination programs. These efforts include changes in malaria diagnosis, i.e. not only diagnosing patients with clinical symptoms of the disease, but also those at risk of malaria [7, 8].

Asymptomatic malaria was found in malaria patients who presented no clinical symptoms of fever during initial inspection but showed the presence of *Plasmodium* sp. on microscopic examination. The persistence of gametocytes in the blood of patients can be a source of new transmission in subsequent malaria infection (gametocyte carrier). Late treatment of asymptomatic malaria detected in patients will increase the chances of transmission of malaria infection and hinder the achievement of malaria elimination [2].

In the early course of the disease, asymptomatic malaria does not present any clinical symptoms, but when antiparasite immunity failed to suppress parasite density, clinical symptoms will appear later [2]. Clinical symptoms are expected to appear two to four weeks later [9, 10]. Dysregulation of the immune system causes asymptomatic malaria, and this will turn into symptomatic malaria and will not be detectable by microscopic (submicroscopic malaria) [11, 12].

The epidemiology of asymptomatic malaria is different in each region and depends on the endemicity. The prevalence of asymptomatic malaria is influenced by many factors, such as age, the density of parasites and insect bites [13]. Asymptomatic malaria may also occur in patients with chronic malaria infection [14]. The biggest challenge today is the accuracy of a diagnostic tool for detecting asymptomatic malaria [2], especially in primary health care.

RDTs, which use the principle of immunochromatography to detect antigens, have been increasingly used as a substitute for microscopic methods, as these are not always readily available in PHC facilities. The efficacy of RDTs, however, may be affected by the target antigen, the number of malaria cases, the density of parasites and the prevalence of parasite species in each region [15]. In rural areas, RDTs are only being used for patients reporting with fever, which is primarily due to certain logistical constraints. This research has shown that 32% of malaria cases were diagnosed in asymptomatic patients, leading to the conclusion that they would not have been detected if the RDTs had only been performed on patients with fever. It can be concluded that the malaria elimination program can only be successful when asymptomatic malaria has been eradicated [16].

Conclusions

Asymptomatic malaria could be detected when RDTs were performed in all inhabitants, not only used in symptomatic patients. Periodic mass blood surveys are necessary to identify asymptomatic malaria cases in hypoendemic areas.

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